

A look into genome of *Ancylostoma* ceylanicum a parasite to human

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Selected paper

genetics

The genome and transcriptome of the zoonotic hookworm *Ancylostoma ceylanicum* identify infection-specific gene families

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Hookworms infect over 400 million people, stunting and impoverishing them¹⁻³. Sequencing hookworm genomes and finding which genes they express during infection should help in devising new drugs or vaccines against hookworms^{4,5}. Unlike other hookworms, Ancylostoma ceylanicum infects both humans and other mammals, providing a laboratory model for hookworm disease6,7. We determined an A. ceylanicum genome sequence of 313 Mb, with transcriptomic data throughout infection showing expression of 30,738 genes. Approximately 900 genes were upregulated during early infection in vivo, including ASPRs, a cryptic subfamily of activation-associated secreted proteins (ASPs)8. Genes downregulated during early infection included ion channels and G protein-coupled receptors; this downregulation was observed in both parasitic and free-living nematodes. Later, at the onset of heavy blood feeding, C-lectin genes were upregulated along with genes for secreted clade V proteins (SCVPs), encoding a previously undescribed protein family. These findings provide new drug and vaccine targets and should help elucidate hookworm pathogenesis.

closely related to the free-living Caenorhabditis elegans than is the free-living Pristionchus pacificus (Fig. 2)^{12–15}. Treatments effective against A. ceylanicum might thus also prove useful against other strongylids, such as Haemonchus contortus, that infect farm animals and depress agricultural productivity¹⁶. Characterizing the genome and transcriptome of A. ceylanicum is a key step toward such comparative analysis.

We assembled an initial *A. ceylanicum* genome sequence of 313 Mb and a scaffold N50 of 668 kb, estimated to cover -95% of the genome, with Illumina sequencing and RNA scaffolding^{17,18} (**Supplementary Tables 1–3**). The genome size was comparable to those of *Ancylostoma caninum* (347 Mb)¹⁹ and *H. contortus* (320–370 Mb)^{20,21} but larger than those of *N. americanus*, *C. elegans* and *P. pacificus* (100– 244 Mb)^{22–24}. We found that 40.5% of the genomic DNA was repetitive, twice as much as in *N. americanus*, *C. elegans* or *P. pacificus* (17–24%). We predicted 26,966 protein-coding genes²⁵ with products of ≥100 residues (**Supplementary Table 4**). We also predicted 10,050 genes with products of 30–99 residues, to uncover smaller proteins that might aid in parasitism²⁶. With RNA sequencing (RNA-seq), we detected expression of 23,855 (88.5%) and 6,883 (68.5%) of these genes, respectively (**Fig. 3**).

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A. ceylanicum Taxonomy

- <u>K:</u> Animalia
 - <u>P:</u>Nematoda
 - <u>C:</u>Chromadorea
 - <u>O:</u>Rhabditida
 - <u>F:</u>Ancylostomatidae
 - <u>G:</u> Ancylostoma
 - <u>S:</u> A. ceylanicum

A. ceylanicum: History & Discovery

- Discovered in 1911 by Arthur Looss.
- Studying bilharzia in Egypt.
- A. braziliense and *A. ceylanicum* was Considered synonymous due to the apparent similarities in almost all respects.





A. ceylanicum: Physical characteristic

- parasitic roundworm .
- about 6–10 mm long.
- Life span from 1-15 years
- Have a mouth like other hookworm with cutting plate with a sharp dorsal end
- Female :tapered narrow posterior end
- Male: have a feathery posterior end.







Ancylostoma duodenale Ancylostoma ceylanicum

Necator americanus



A. ceylanicum: Geographic distribution



A. ceylanicum: infection information

- A. ceylanicum enter to the host
 by burrowing into skin.
- Pass with the bloodstream until it leads to the small intestine.
- In small intestine it feed on the blood and causing animea.
- accompanied significant loss in body weight.
- It may cause iron deficiency, hypoalbuminemia, in addition to animea.

A. ceylanicum: Parasitism

• A. ceylanicum dose not have an intimidate host. Therefore human can be primary host.

• Can be seen in the skin of the host.



A. ceylanicum: Sequencing sample info

 All specimens of Ancylostoma ceylanicum were collected from Golden Syrian hamsters.

The Aroian laboratory strain of A. ceylanicum used in this work has been designated HY135



A. ceylanicum : Sequencing method

Illumina sequencing method



A. cevlanicum : Genome assembly

Supplementary Table 2: Characteristics of genome and cDNA assemblies.

	Genome assemblies					cDNA
	A. ceylanicum	C. elegans	P. pacificus	N. americanus	H. contortus	assembly (from RNA-seq)
Total nt:	313,110,363	100,286,401	172,494,865	244,075,060	369,846,877	64,318,273
Scaffolds:	1,737	7	18,083	11,864	23,860	332,724
Contigs:	32,171	7	33,305	65,213	65,523	332,724
ACGT nt:	300,908,004	100,286,401	153,192,245	208,173,610	346,042,478	64,318,273
N-res. nt:	12,202,359	0	19,302,620	35,901,450	23,804,399	0
% non-N:	96.1	100.0	88.8	85.3	93.6	100.0
% GC:	43.4	35.4	42.8	40.2	43.1	44.6
Scaffold N50 nt:	668,412.0	17,493,829.0	1,244,534.0	211,861.0	83,287.0	294.0
Scaffold N90 nt:	117,063.0	13,783,801.0	85,679.0	29,168.0	11,529.0	78.0
Scaf. max. nt:	4,802,298	20,924,180	5,268,024	1,890,151	947,606	10,003
Scaf. min. nt:	509	13,794	47	201	101	53
Contig N50:	18,451.0	17,493,829.0	18,131.0	5,429.0	20,808.0	294.0
Contig N90:	4,900.0	13,783,801.0	2,062.0	1,285.0	4,267.0	78.0
Contig max. nt:	125,366	20,924,180	163,374	62,795	135,785	10,003
Contig min. nt:	1	13,794	1	25	1	53

Statistics are shown for the *A. ceylanicum* genome and cDNA assemblies, along with genome assemblies from the closely related nematodes *C. elegans*, *P. pacificus*, *N. americanus*, and *H. contortus*. Assemblies from related nematodes were from WormBase release WS242. For *H. contortus*, we used the genome assembly of Laing et al.⁸²².

-Genomic assembly

- Contigs= 32 Kb
- Genome size = 347 Mb
- Scaffolds = 1 kb
- Scaffolds N50 = 668 Kb

-cDNA assembly

- Contigs= 332 kb
- Genomic size=64 Mb
- Scaffolds= 332 kb
- Scaffolds N50=294

A. ceylanicum : Interesting genome outcome

- About 40.5% of the genomic DNA was repetitive.
- The GC% was about 43.4% for genomic DNA while 44.6 for cDNA.

Questions

1. What is the intermediate host of *A. ceylanicum*?

2. what was the sequencing method that was used and give a brief description about the steps?

Thank You for Listening