



**Research Topics and Seminar
(501)
Dr. Hassan Al-Haddad**

In vitro embryo production in camel (*Camelus dromedarius*) from in vitro matured oocytes fertilized with epididymal spermatozoa stored at 4 °C

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In vitro embryo production in camel (*Camelus dromedarius*) from in vitro matured oocytes fertilized with epididymal spermatozoa stored at 4 °C

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Abstract

Experiments were conducted to study the effect of storing epididymal spermatozoa, in tris–tes- and tris–lactose egg yolk extenders, on their fertilizing ability and subsequent in vitro embryo development. Ovaries and testes were collected from a local slaughterhouse in normal saline solution (NSS) at 37 °C and on ice (0–1 °C), respectively. Cumulus oocyte complexes (COCs) aspirated from the follicles were randomly distributed to 4-well culture plates (20–25 COCs/well) containing 500 µL of maturation medium and cultured at 38.5 °C in an atmosphere of 5% CO₂ in air for 36 h. Spermatozoa were collected from the cauda epididymides in syringes containing 2–3 mL of either tris–tes- or tris–lactose egg yolk extender. They were cooled down slowly and stored at refrigeration (4 °C) temperature. The spermatozoa were evaluated for motility and used for IVF of IVM oocytes on the day of collection and after 2, 4, 6 and 8 days of storage. On the day of IVF, spermatozoa were prepared by the swim up technique and both spermatozoa and oocytes were co-incubated at 38.5 °C in a humidified atmosphere of 5% CO₂ in air for 15–16 h. Presumptive zygotes were either fixed and stained with Hoechst 33342 for evaluation of fertilization or were cultured in 500 µL of the culture medium at 38.5 °C in an atmosphere of 5% CO₂, 5% O₂ and 90% N₂ in air. There was no significant difference ($P > 0.05$) in the proportion of oocytes fertilized with spermatozoa stored in either of the two extenders for up to 8 days. The proportion of oocytes that cleaved (43–60%) and those that developed to blastocysts (14–21%) did not show any difference ($P > 0.05$) either, when spermatozoa from different days of storage were used. First cleavage was observed as early as 16 h after IVF, early blastocysts had developed by day 4, expanded blastocysts after day 5 and hatching of blastocysts started after day 6 of culture.

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Outline

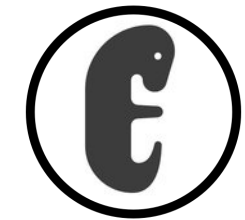
- Introduction (general information about camel and Breeding)
- Objective
- Method
- Result
- Discussion
- Conclusion



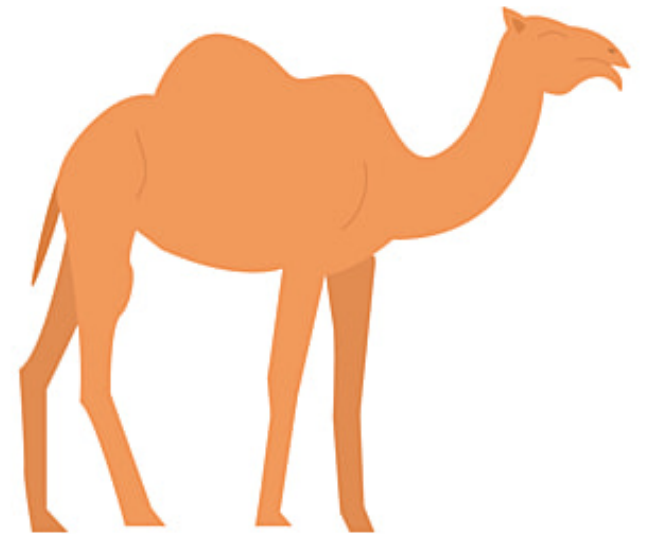
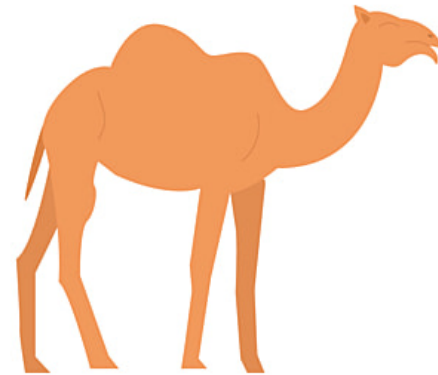
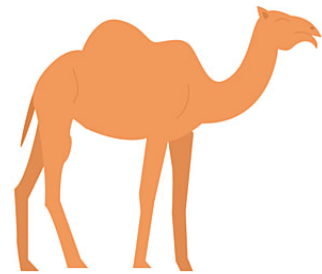
Introduction

Camel

- Scientific name of the camel is *Camelus*.
- Camel can live for 40 years.

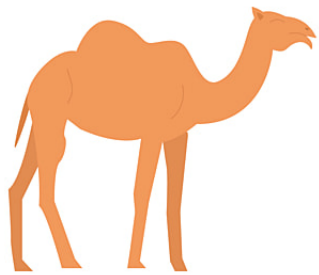


EMBRYO



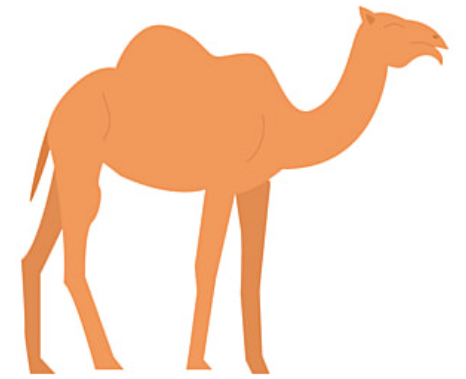
Sexual maturity

- Male: at 5 years.
- Female: at 3 to 4 years



IMMATURE

Years



MATURE

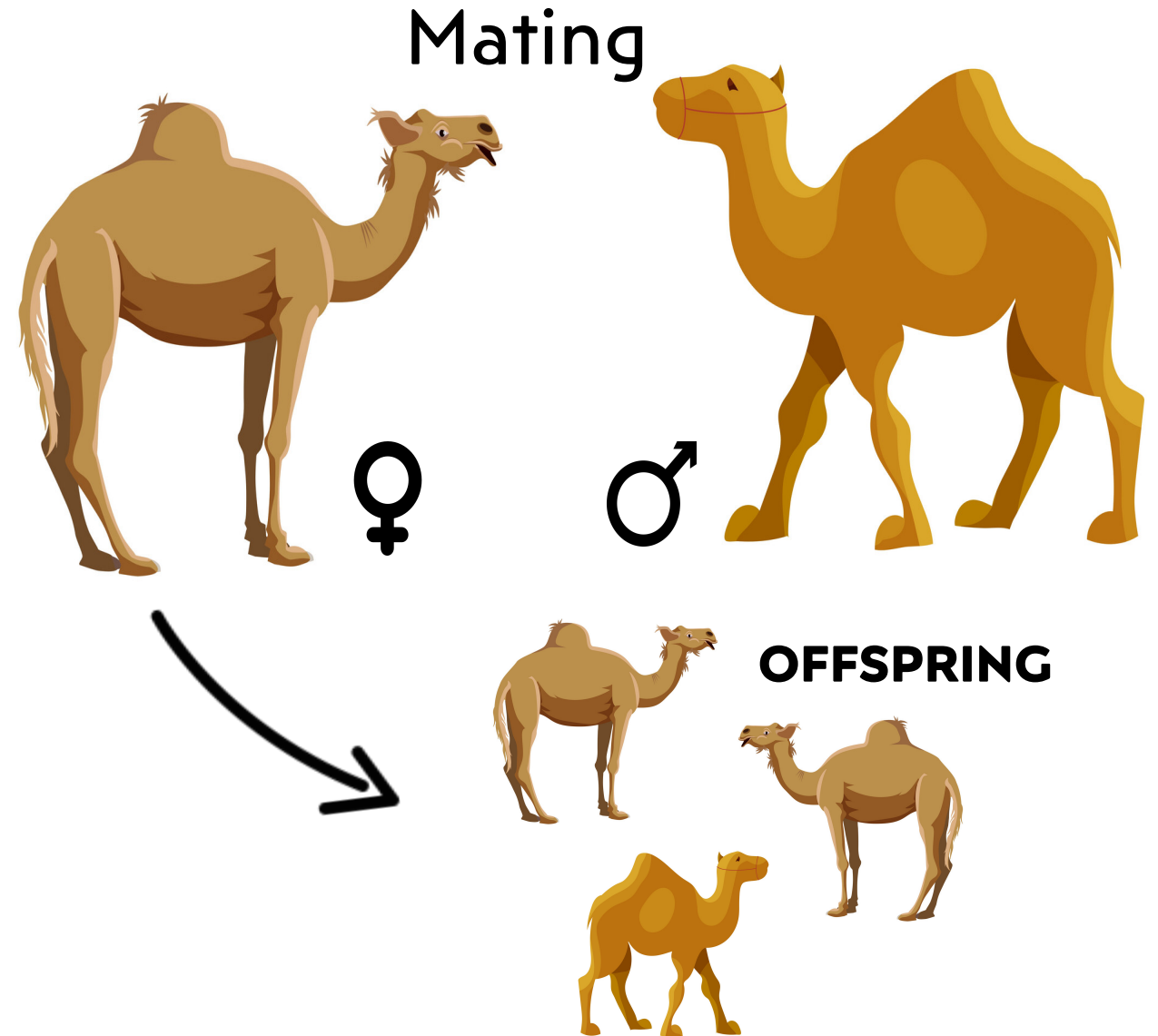
Breeding season

Sudan	between March and August
Egypt	December to April
India	November to February



Reproduction

- It happens when male camel mates female camel.
- This is the natural way.



Camel pregnancy

- Pregnancy is characterized by the presence of a large and well developed CL (CORPUS LUTEUM) which is maintained throughout pregnancy.
- The gestation length takes from 315 to 440 days.
- Birth weight of dromedary calves varies from 19kg to 52 kg

In vitro fertilization

- It is the artificial way in which it happened in the lab.

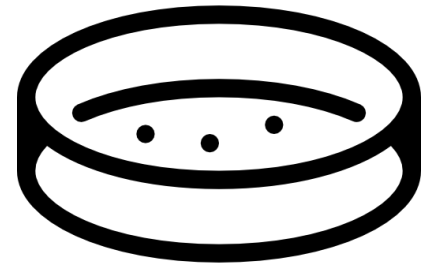


Lab



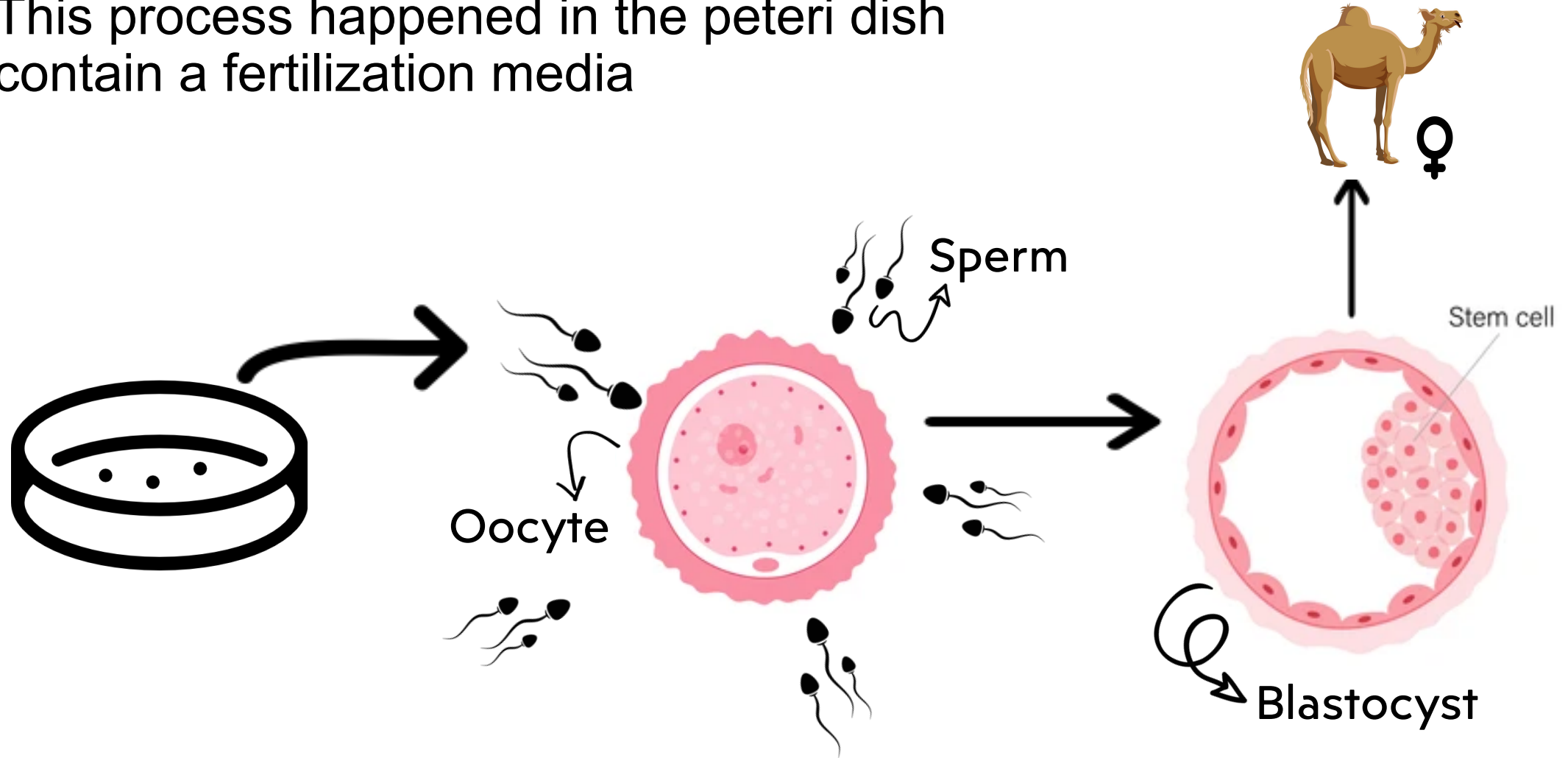
Embryologist

Petri dish



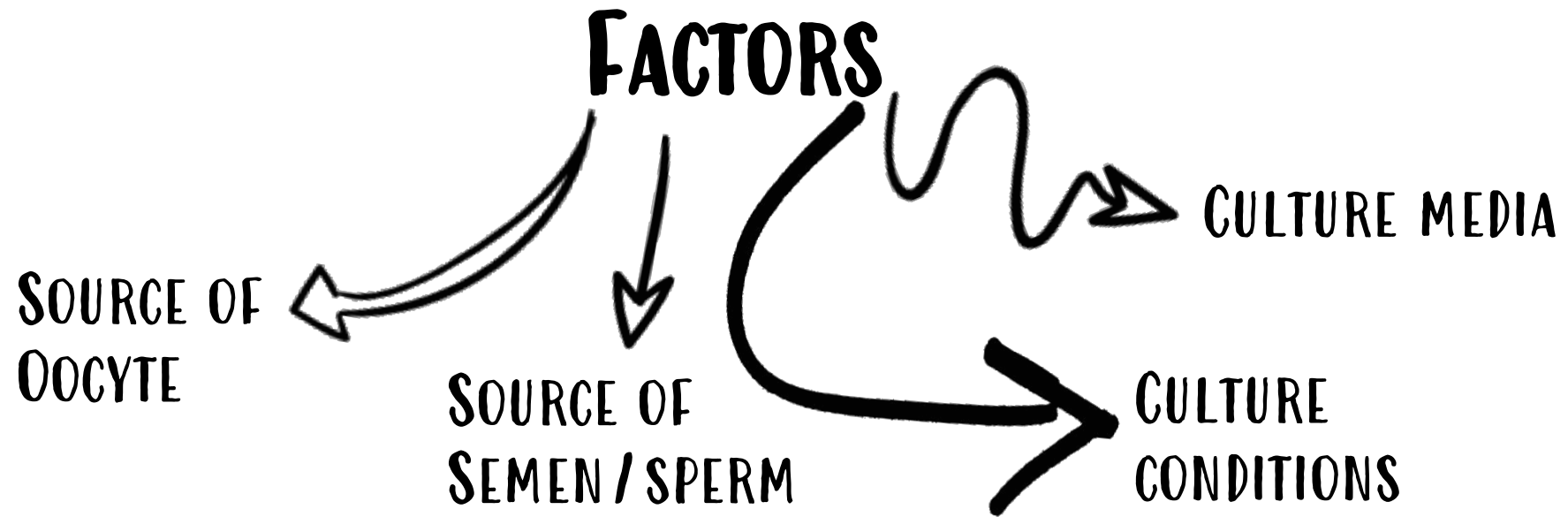
In vitro fertilization

- This process happened in the petri dish contain a fertilization media



In vitro fertilization

- There are many factors affecting the process





Objective

Objective

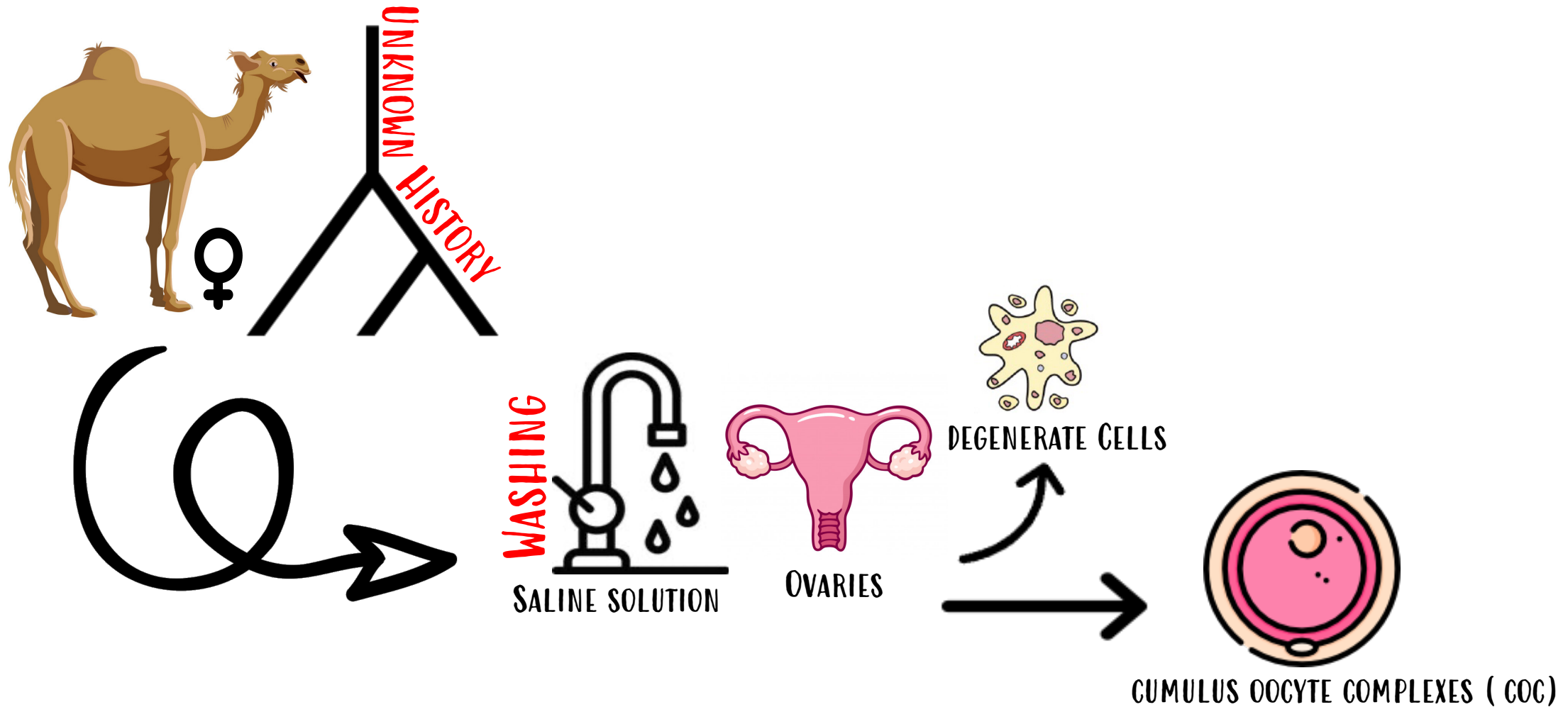
- To determine the effect of storing epididymal spermatozoa on fertilizing ability of the sperm.



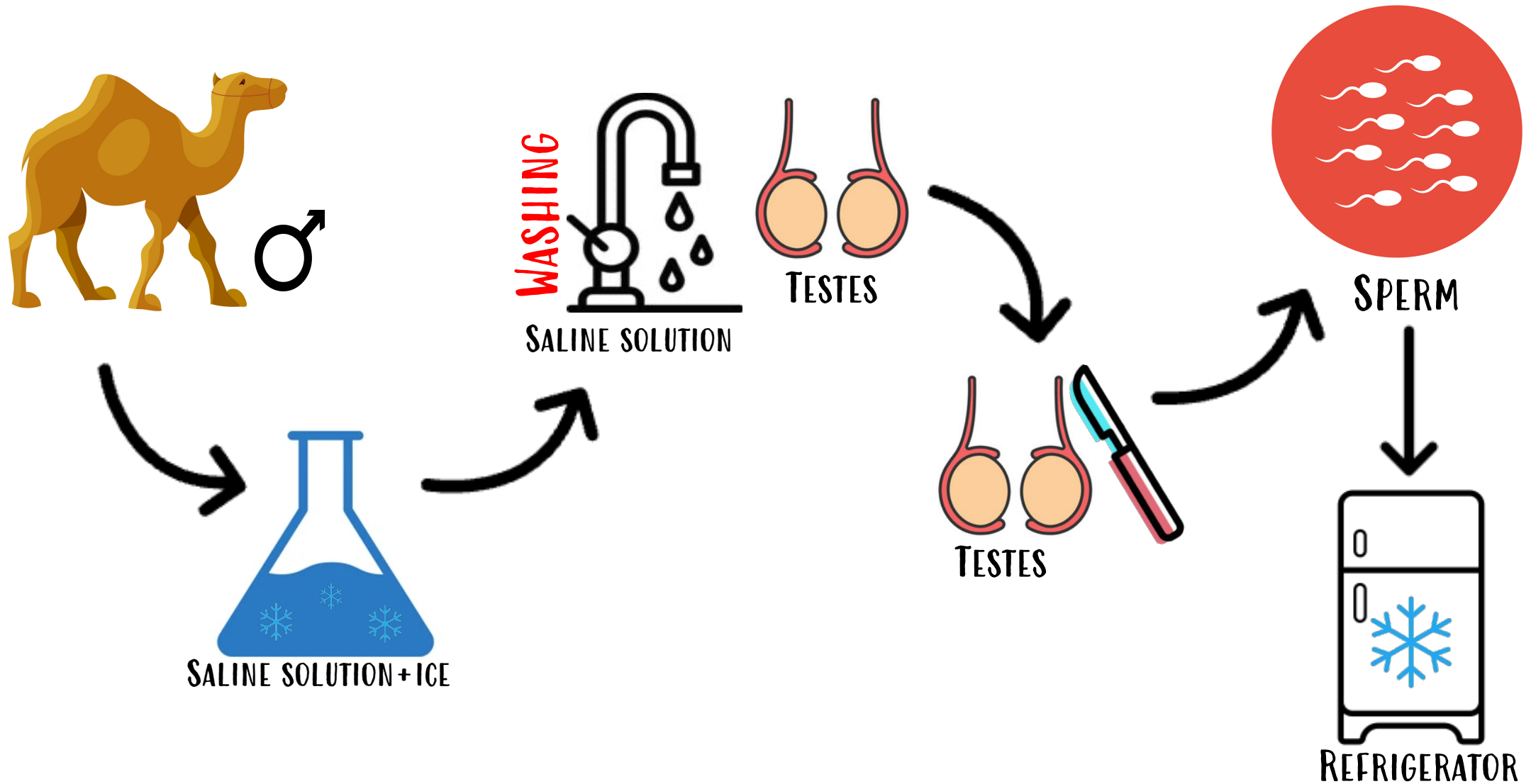
Material and methods



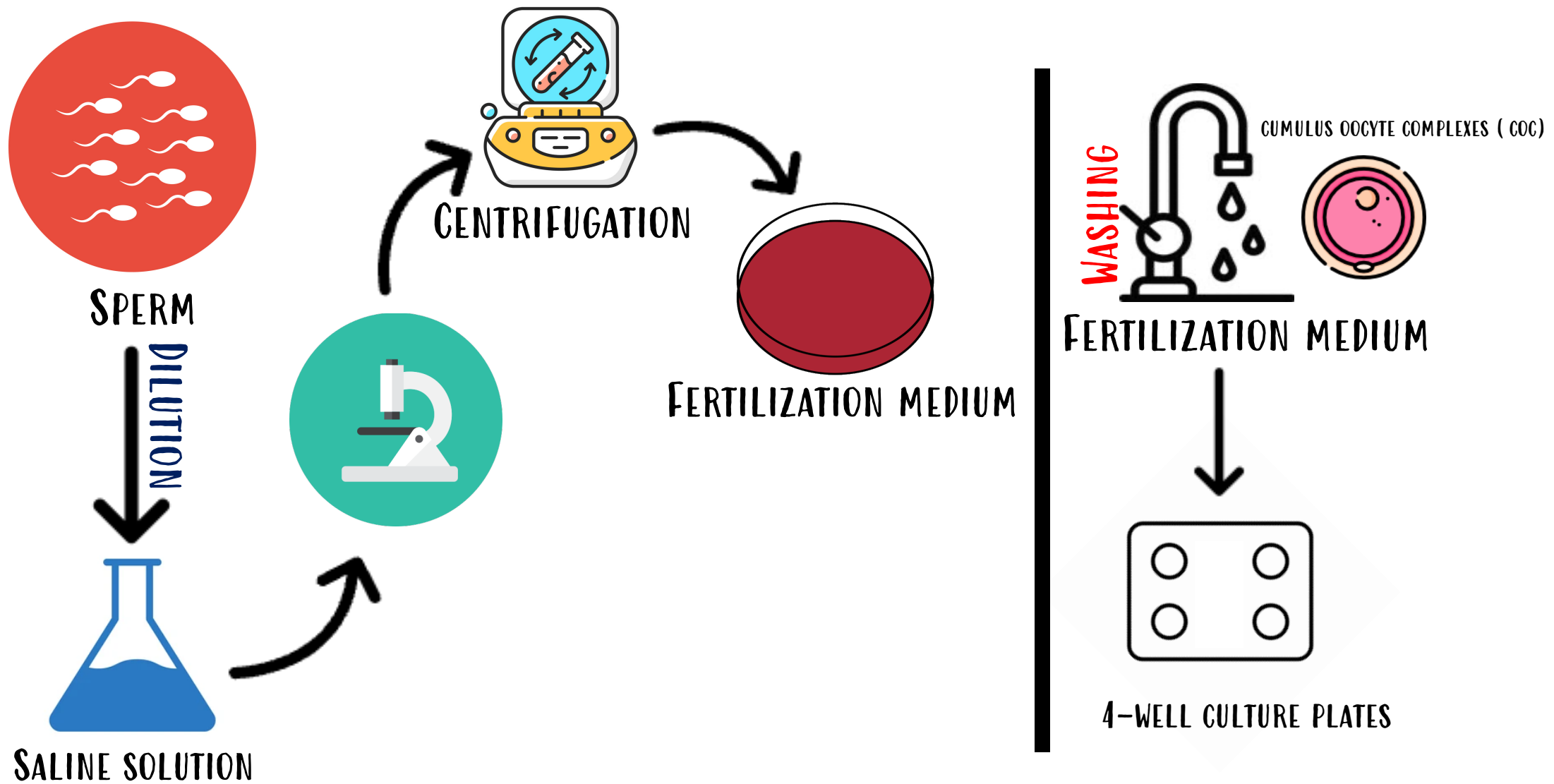
Collection of COC



Collection and storage of semen

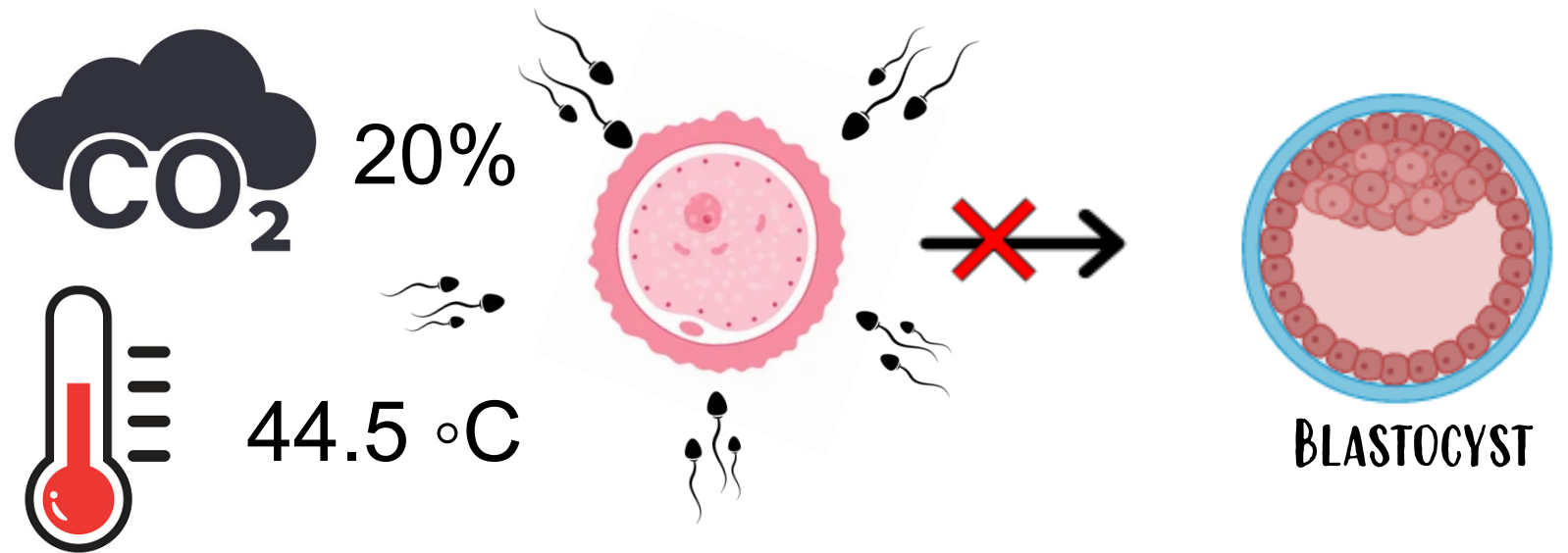
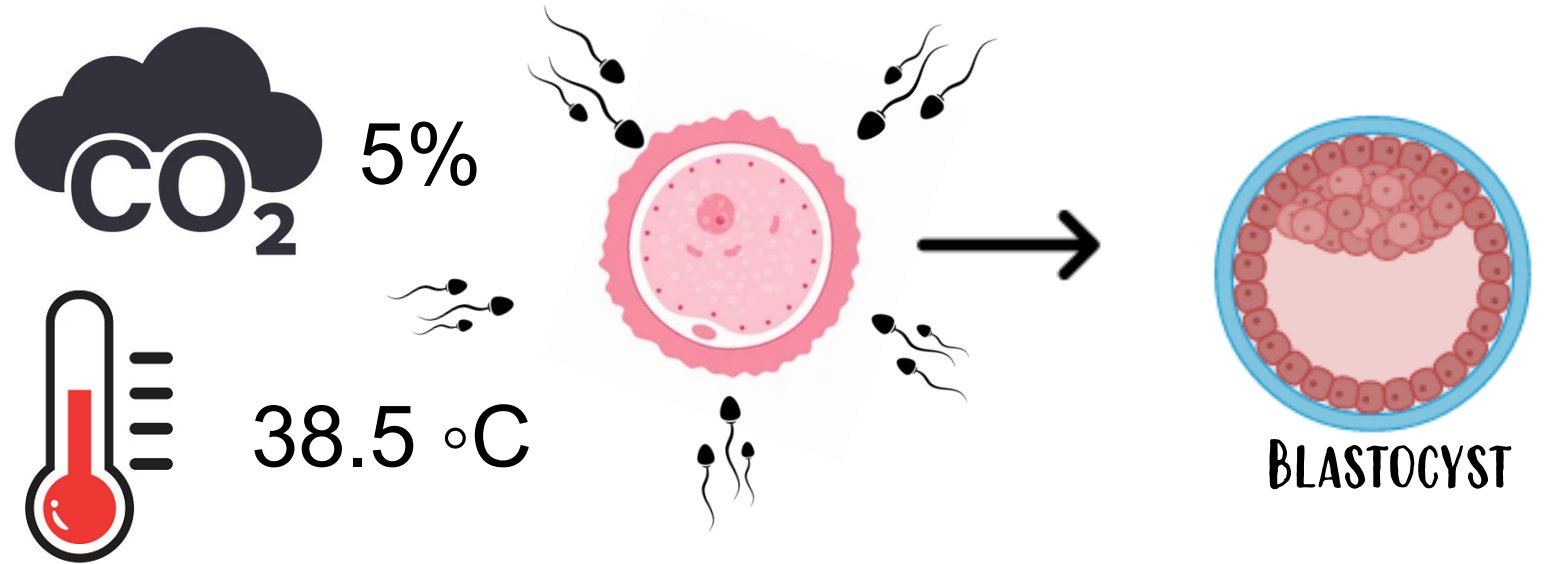


In vitro fertilization

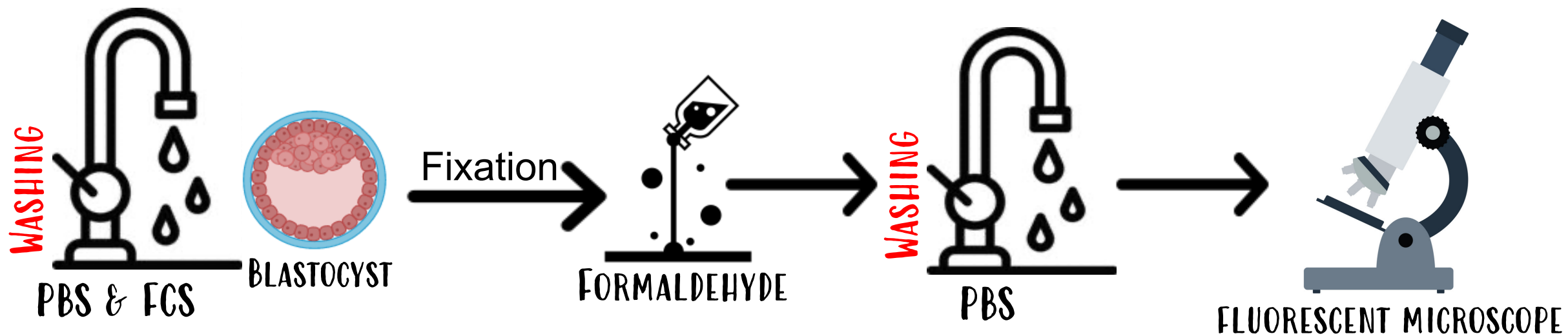


Conditions:

- There are conditions that enhance the fertilization process.



Evaluation of fertilized oocyte



Statistical Analysis

- Some statistical tests were done:
 - I. T-test
 - II. Chi- square test



Result



In vitro fertilization

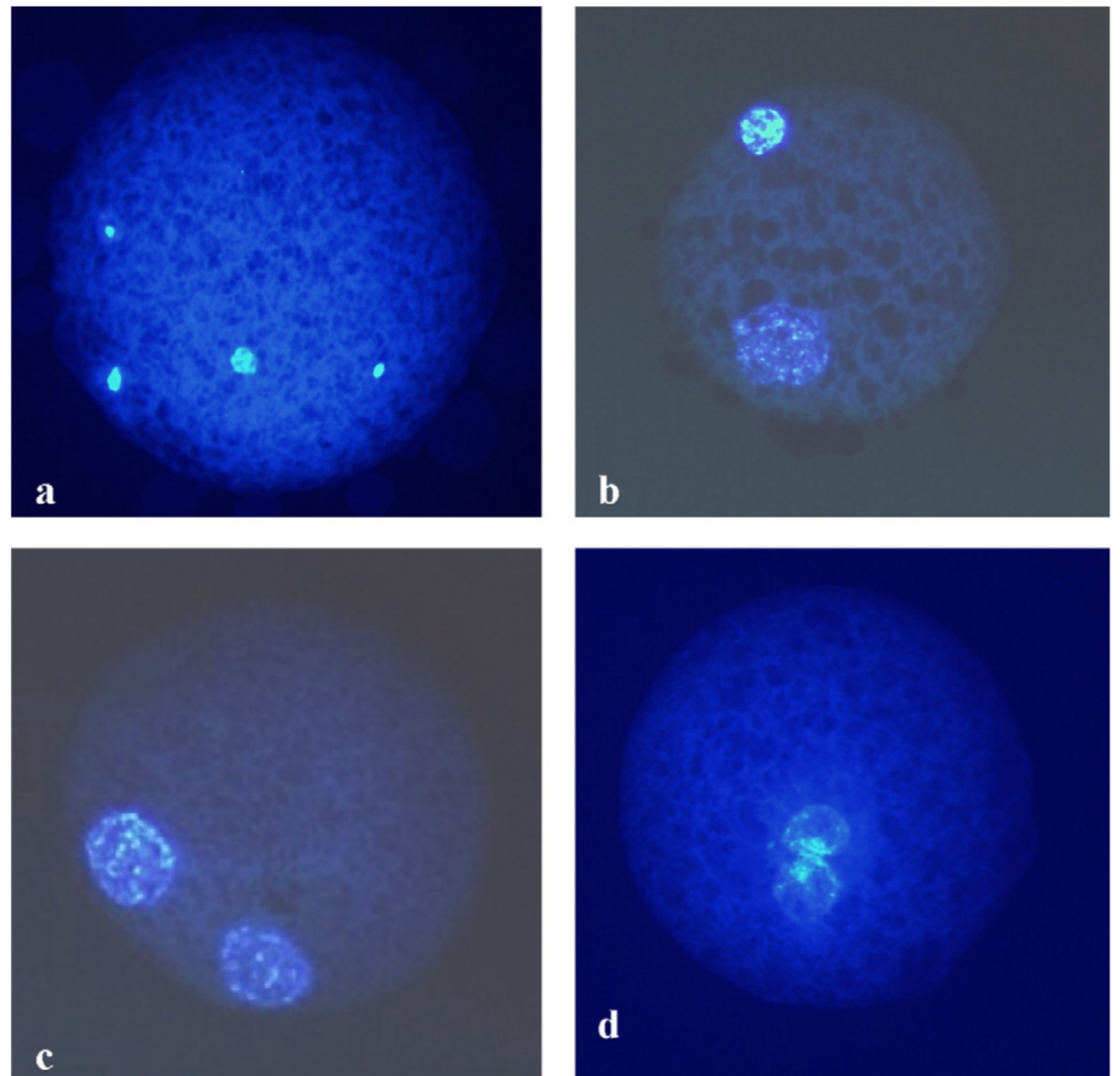
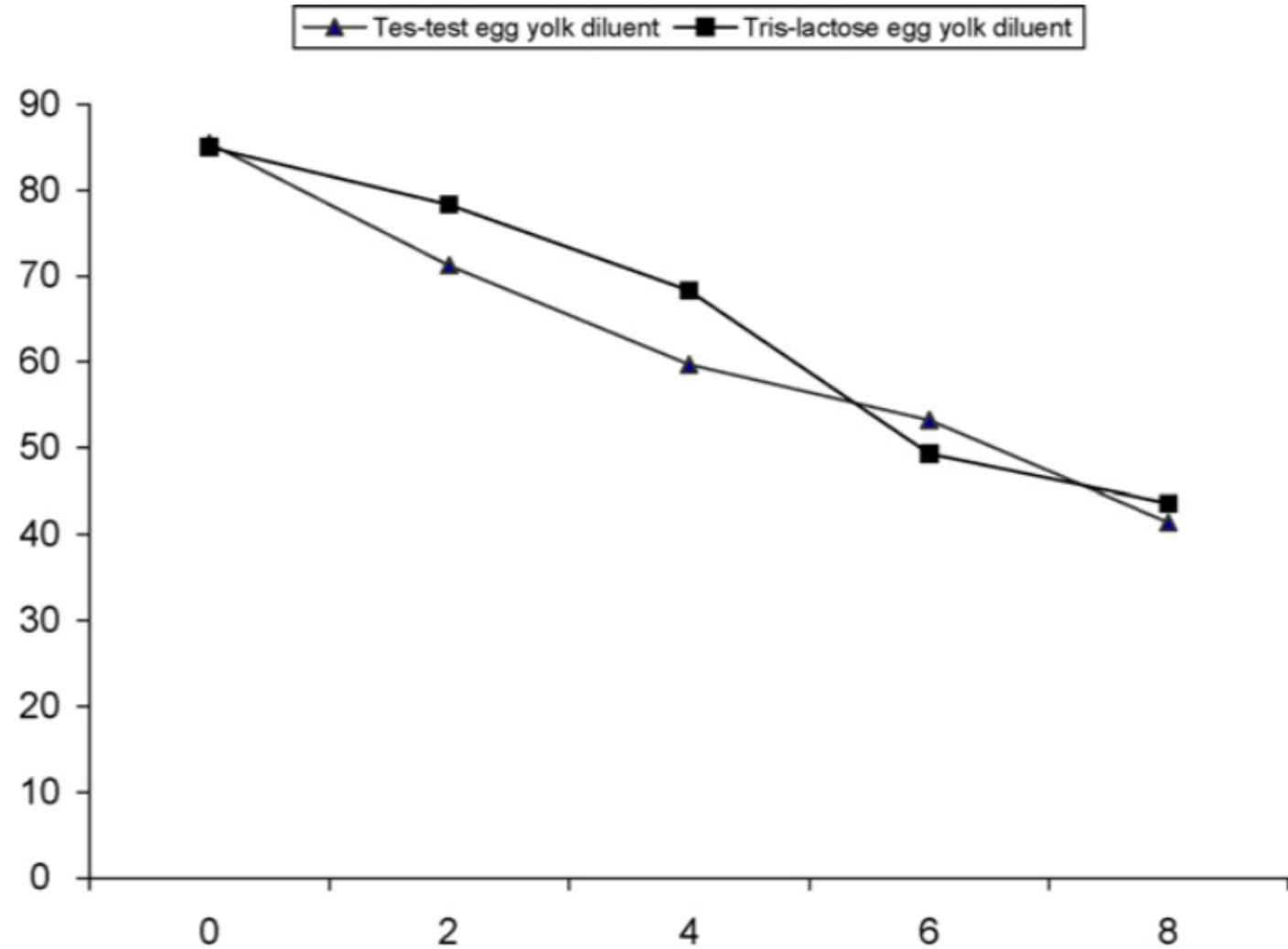


Fig. 1. In vitro fertilized oocytes: (a) with a sperm in ooplasm and visible first and second polar bodies; (b) with a female pronucleus and condensation of sperm chromatin; (c) male and female pronuclei and (d) a syngamous male and female pronuclei.

Sperm motility

- There was slow loss of spermatozoa motility.
- There was no difference in motility between Two types of sperm

N.A. Wani / Animal Reproduction Science 111 (2009) 69–79



Fertilization rate

Table 1

Fertilization rates of the in vitro matured oocytes with the epididymal spermatozoa stored in tris–tes- and tris–lactose egg yolk extenders at 4 °C up to 8 days

Day of semen storage	Extender used	Total oocytes (%)			
		Inseminated	Fertilized normal	Polyspermic	Total
0	Tris–tes	102	58 (56.9)	3 (2.9)	61 (59.8)
	Tris–lactose	92	59 (64.1)	3 (3.2)	62 (67.4)
2	Tris–tes	96	58 (60.4)	4 (4.2)	62 (64.6)
	Tris–lactose	90	60 (66.7)	3 (3.3)	63 (70.0)
4	Tris–tes	93	50 (53.8)	7 (7.5)	57 (61.3)
	Tris–lactose	92	57 (61.9)	5 (5.4)	63 (68.5)
6	Tris–tes	95	49 (51.6)	5 (5.3)	54 (56.8)
	Tris–lactose	90	55 (61.1)	3 (3.3)	58 (64.4)
8	Tris–tes	89	51 (57.3)	7 (7.9)	57 (64.0)
	Tris–lactose	97	59 (60.8)	2 (3.3)	61 (62.9)

Development of matured oocyte

Table 2

Development of in vitro matured oocytes fertilized with epididymal spermatozoa stored in tris–tes- and tris–lactose egg yolk extenders at 4 °C up to 8 days

Day of semen storage	Extender for semen storage	Total oocytes inseminated	Cleaved (%)	Blastocyst			Hatched blastocysts (%) ^c
				No	(%) ^a	(%) ^b	
0	Tris–tes	119	56 (47)	11	9	20	4 (36)
	Tris–lactose	115	61 (53)	13	11	21	5 (38)
2	Tris–tes	120	66 (55)	14	12	21	5 (36)
	Tris–lactose	117	70 (60)	15	13	21	8 (40)
4	Tris–tes	124	65 (52)	10	8	15	3 (30)
	Tris–lactose	121	70 (58)	14	12	20	5 (36)
6	Tris–tes	113	52 (46)	8	7	15	2 (25)
	Tris–lactose	109	60 (55)	11	10	18	3 (27)
8	Tris–tes	123	57 (46)	8	6	14	2 (25)
	Tris–lactose	129	56 (43)	10	8	18	4 (33)

^a Percent from total oocytes.

^b Percent from cleaved embryos.

^c Percent from blastocysts.

Blastocyst development

- First cleavage was seen after 16 h from IVF.
- Early blastocysts had developed by day 4.
- Expanded blastocysts after day 5 and hatching of blastocysts started after day 6 of culture.

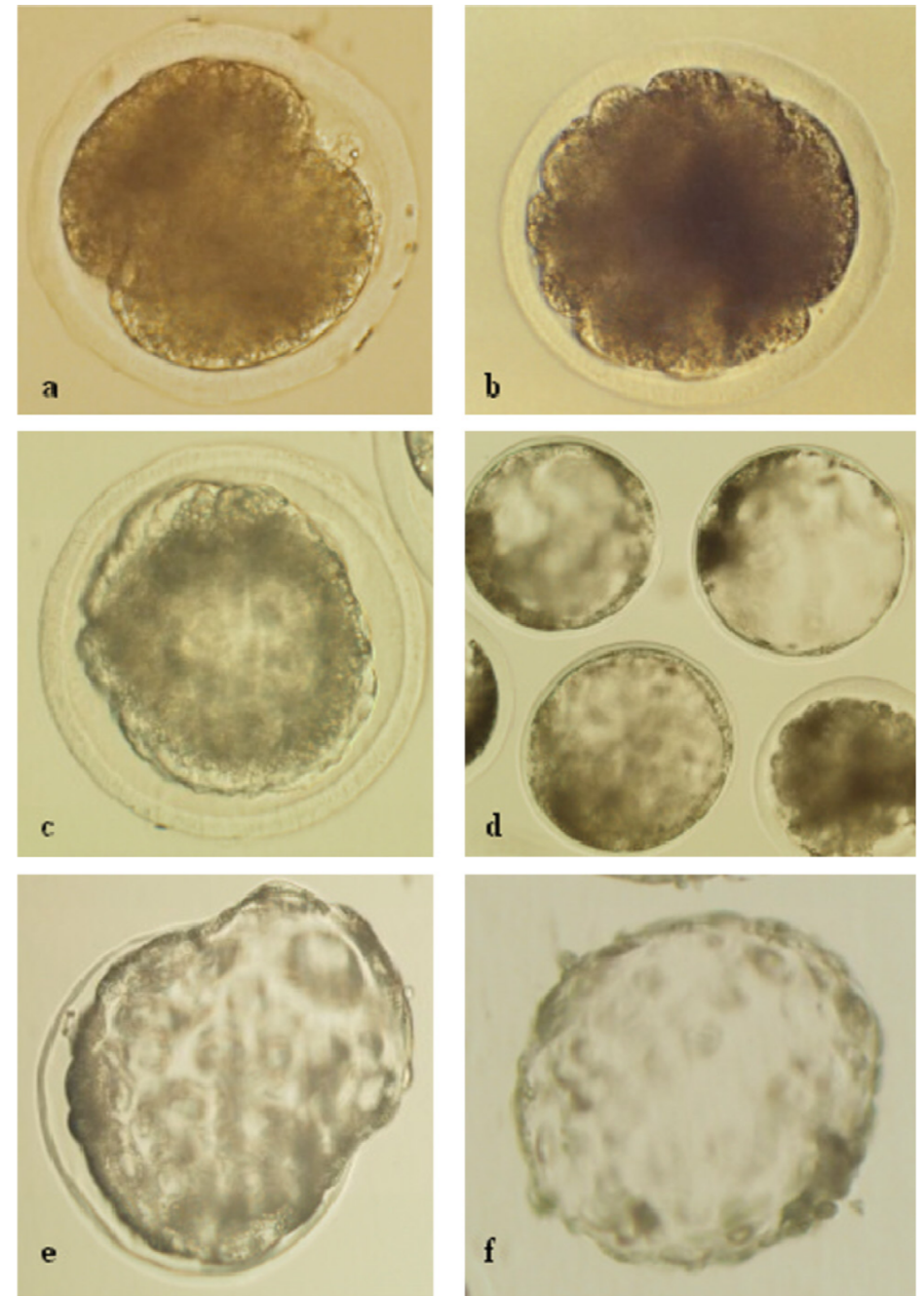


Fig. 3. In vitro embryo production in dromedary camel: (a) a 2-cell embryo; (b) a morula; (c) an early blastocyst; (d) expanded blastocysts; (e) a hatching blastocyst and (f) a hatched blastocyst.

Discussion

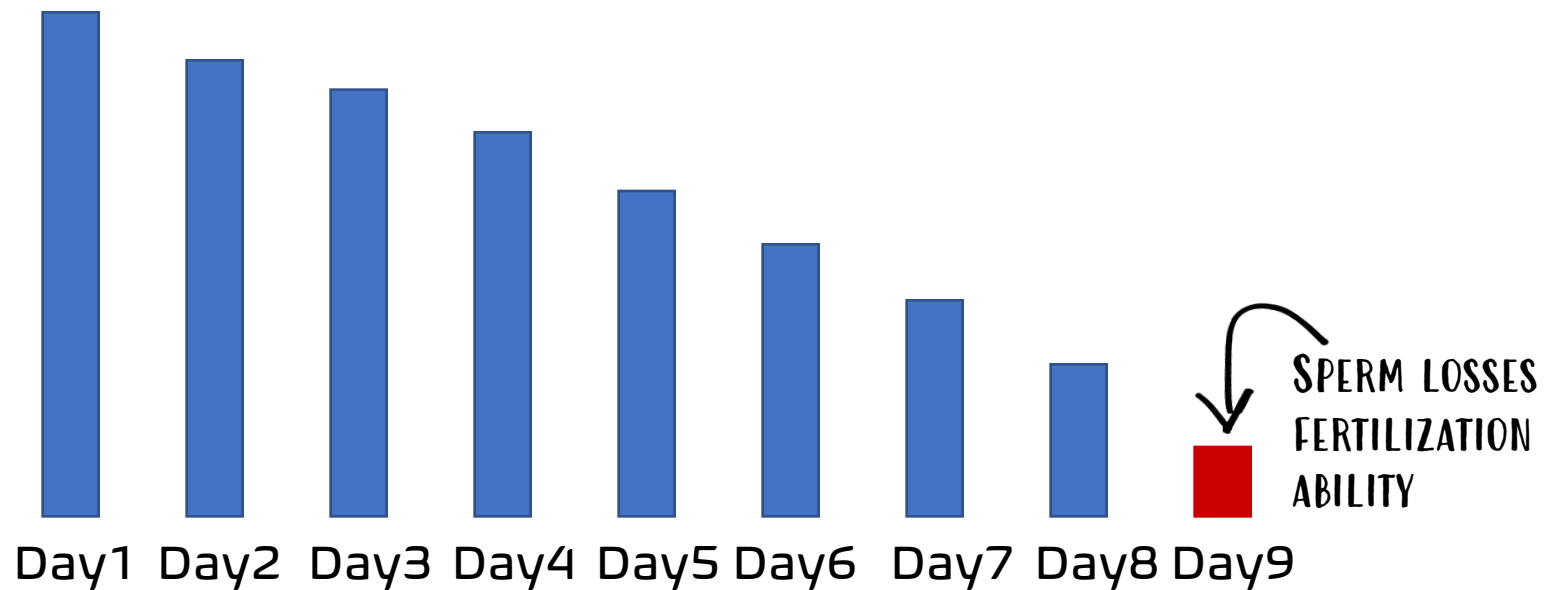


Fertilization ability

- Epididymal spermatozoa was able to fertilize mature oocytes in vitro after storage at 4 °C in tris–lactose and tris–tes extenders for at least 8 days.



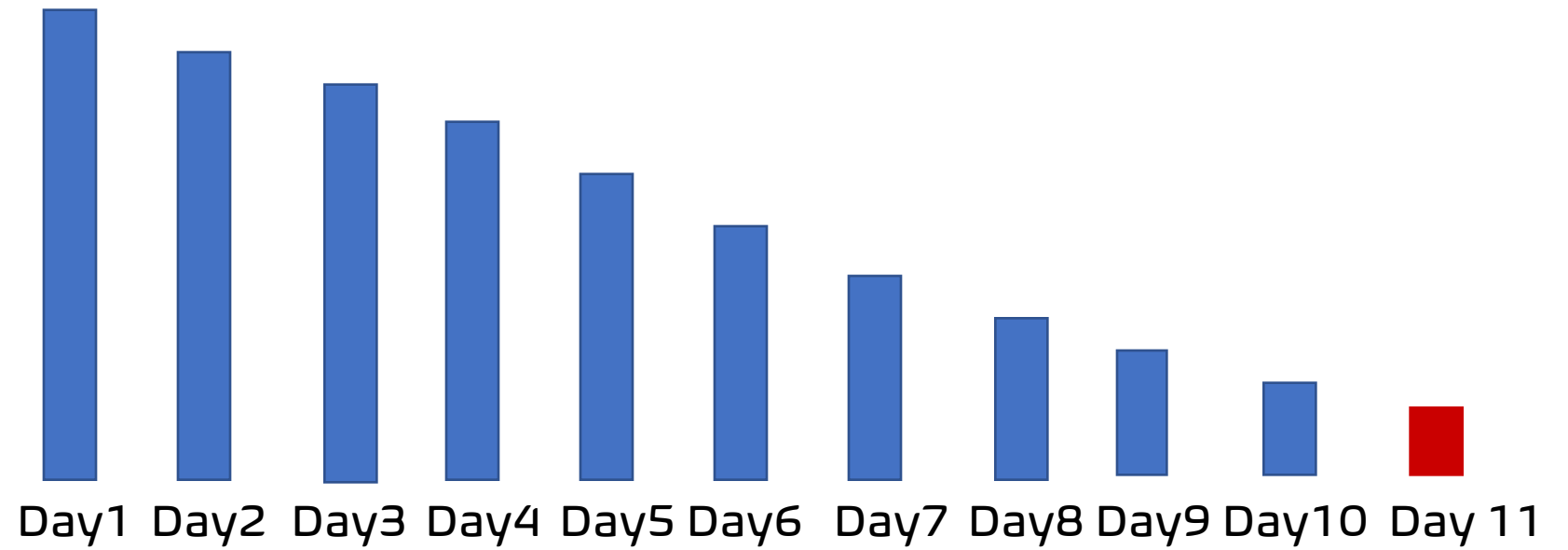
Camel



Fertilization ability



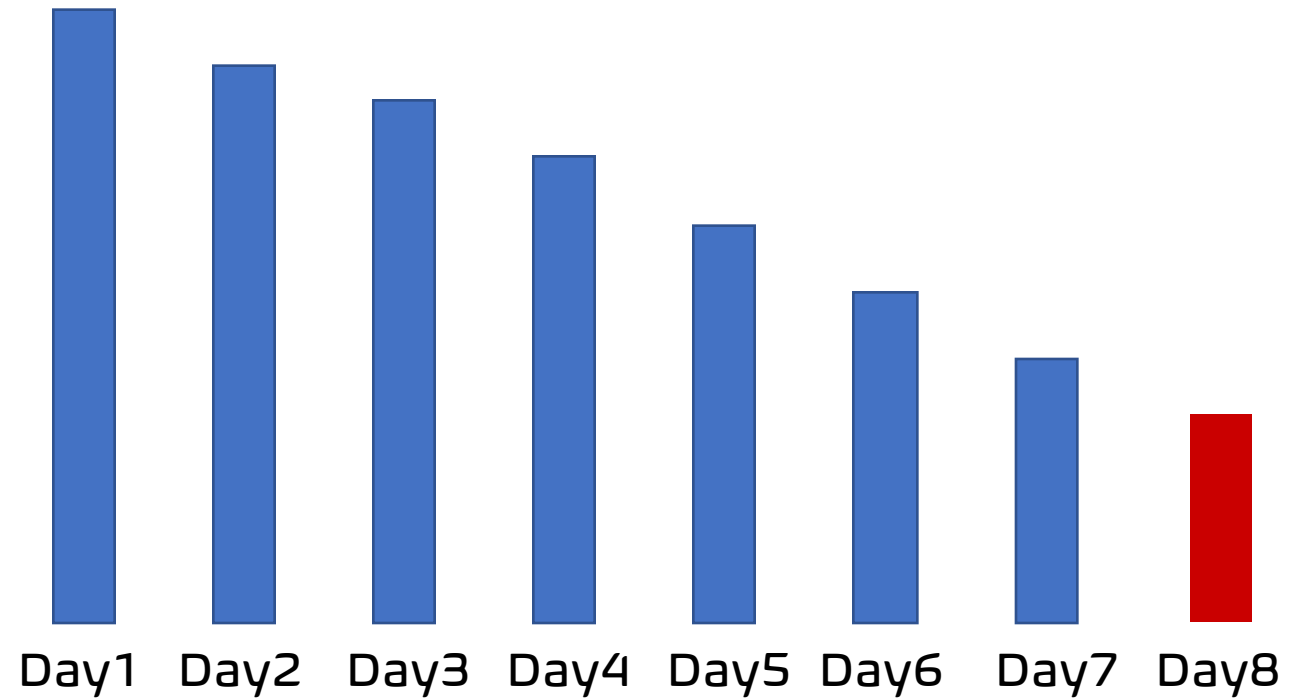
Bull



Fertilization ability



Cat

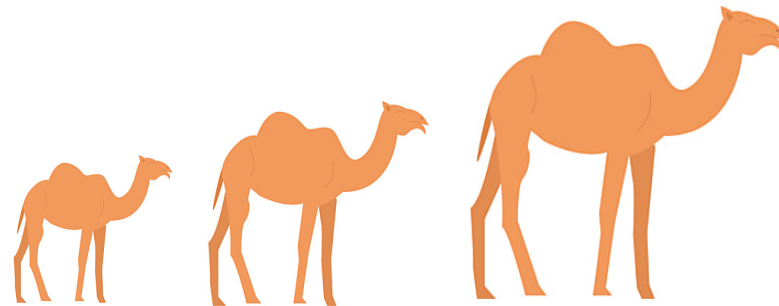


Fertilization rate

- The fertilization rate of epididymal spermatozoa may differ due to:



Site of sperm collection



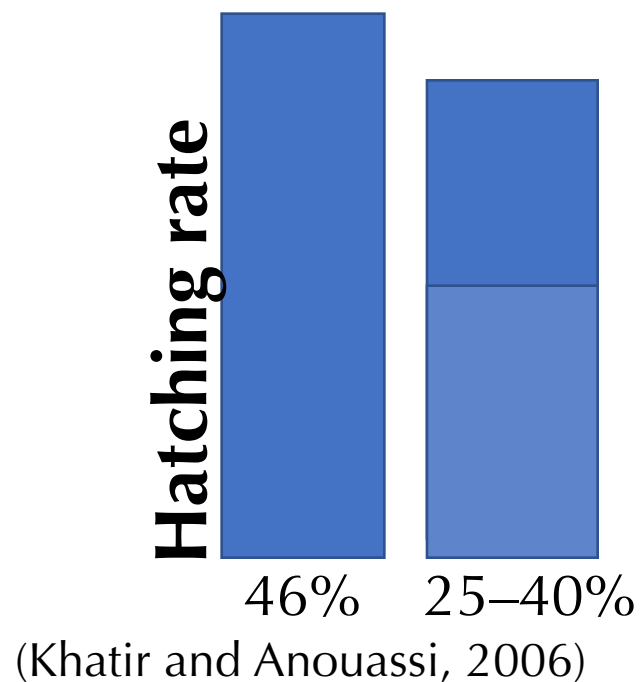
age of the animals



methods of semen preparation

Hatching rate

- Hatching rate was slightly lower than the study published before.
- The reasons for this variability might be the supplements in embryo culture medium.



Embryo development

- The chronology of embryo development in dromedary camels is faster than in other species.
- In cattle the blastocyst is seen after 6–7 days of culture and hatches after day 7 while in llamas the blastocysts hatch on day 7 after IVF.

16 h post IVF	first cleavage was observed
Day 6	first hatched blastocysts



Conclusion



Conclusion

- The dromedary epididymal spermatozoa survive in storage for at least 8 days in tris–lactose and tris–tes egg yolk diluents at 4°C.
- These spermatozoa maintain their fertilizing ability and may be suitable for use in IVF.



Reference:

- Wani, N., 2009. In vitro embryo production in camel (*Camelus dromedarius*) from in vitro matured oocytes fertilized with epididymal spermatozoa stored at 4°C. *Animal Reproduction Science*, 111(1), pp.69-79.
- Eiwishy, A., 1987. Reproduction in the female dromedary (*Camelus dromedarius*): A review. *Animal Reproduction Science*, 15(3-4), pp.273-297.